# Causes and consequences of the variation in the number of ovarian follicles in cattle

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# Summary

In cattle we have noted that the antral follicle count (AFC, follicles  $\geq$  3 mm in diameter) varies greatly among animals (from 5 to 50), is repeatable within animals, and is highly correlated with the total number of healthy follicles in ovaries. Also, animals with low AFC have higher serum concentrations of FSH and LH, but lower concentrations of Anti-Mullerian Hormone, progesterone and androgens than animals with high AFC. We have investigated the effect of maternal environment during gestation on their offspring AFC by restricting maternal nutrition to 60% of maintenance requirements (compared with 100% in controls) during the first third of gestation. Calves born to nutritionally restricted mothers had 60% lower AFC compared with calves born to mothers fed control diets. In other studies we have evidence to indicate that fertility may be compromised in animals with low AFC due to effects on oocytes, progesterone and the endometrium compared with animals with high AFC. To examine this directly we assessed AFC in post-partum dairy cows and found that cows with a high AFC had higher pregnancy rates, shorter calving to conception intervals and received fewer services during the breeding season compared with cows with a low AFC. In addition, the high variation in follicle numbers in adults may not only be reflective of reproductive disorders and suboptimal fertility, but there is evidence to indicate that it may be associated with alterations in the function of other non-reproductive systems (e.g. cardiovascular) that may have profound effects on the animal's health and welfare.

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## Introduction

In cattle, growth of ovarian antral follicles from about 300 µm to 5 mm in diameter takes more than 30 days (Lussier et al. 1987). Subsequent follicle growth to 15 to 20 mm in diameter is rapid and occurs in a wave-like pattern over the next 4 to 6 days with two or three waves occurring during the normal course of oestrous cycles in cattle (Evans 2003). The wave-like pattern of follicle growth was first proposed by Rajakoski in 1960 after a large study examining the number of follicles >1 mm in diameter in the ovaries of slaughtered cattle on different days of the cycle (Ireland et al. 2000). Rajakoski's observation, that the numbers of follicles increased and decreased in a wave-like pattern, gave rise to the two follicle wave hypothesis (Rajakoski 1960). Ireland and Roche used intrafollicular concentrations of oestradiol to progesterone to classify antral follicles  $\geq$  6 mm in diameter and proposed the three-wave hypothesis for dominant follicle development in cattle (Ireland & Roche 1982, Ireland & Roche 1983b, Ireland & Roche 1983a, Ireland & Roche 1987). The development of ultrasonography as a tool to noninvasively monitor the growth and regression of individual ovarian follicles repeatedly in the same animal (Pierson & Ginther 1984) firmly established that antral follicles grow in cohorts, in two or three wave-like patterns during oestrous cycles in cattle (Pierson & Ginther 1988, Savio et al. 1988, Sirois & Fortune 1988). This finding was later extended to other reproductive periods in cattle and to other species (Ireland et al. 2000, Evans 2003). Numerous subsequent studies examined the associations among reproductive hormones and follicle growth (Mihm et al. 2002). However, the observations that the number of primordial follicles is highly variable in cattle at birth (Erickson 1966b), that the number of different follicle types vary greatly among adult cattle, and that they reliably ovulate one, or occasionally two, follicles during each oestrous cycle prompted further investigations into the variation in ovarian follicle numbers in cattle.

## Variation in the numbers of ovarian follicles and associated reproductive hormones

#### Follicle numbers

Our research groups have now conducted a number of studies that have systematically counted and catalogued the numbers of antral follicles on different days of the oestrous cycle in both beef and dairy heifers and in post-partum dairy cows. We have established that the numbers of follicles in ovarian follicular waves of the oestrous cycle are highly variable among animals but very highly repeatable within individuals (Burns et al. 2005, Ireland et al. 2007, Ireland et al. 2008, Jimenez-Krassel et al. 2009) (Mossa et al. 2010b). This observation holds true when considering the peak or nadir numbers associated with waves or the mean numbers across all days of the cycle. However, the count must include all the follicles  $\geq 3$  mm in diameter in both ovaries (the antral follicle count or AFC) and explains why many studies in the last 20 years that have focused on follicles  $\geq$  5 or 6 mm in diameter have not noted the high variability of follicle numbers growing during follicular waves among animals nor the remarkably high repeatability of follicle numbers during waves in individuals. For example, we have noted that the AFC in both ovaries during different follicular waves of an oestrous cycle may be consistently lower than 5 during follicular waves in some animals and greater than 50 in others (Burns et al. 2005, Ireland et al. 2007, Mossa et al. 2010b) (Figure 1). Moreover, this high repeatability of follicle numbers during waves persists for at least one year (Burns et al. 2005).

The observation of the variation in AFC raises questions about the possibility for variation in the total numbers of follicles in the ovaries. To address this, age matched heifers (adult cycling animal 12 to 18 months of age) that had high ( $\geq$  25) or low ( $\leq$  15) AFC were identified, and the numbers of all follicles in the ovaries were counted in histological sections (Ireland et al. 2008).

This study showed that the AFC reliably predicted the numbers of morphologically healthy follicles in all classes of the stages of folliculogenesis (Ireland *et al.* 2008) (Figure 2). For example, cattle with a low AFC also had a very low total number of healthy primordial, preantral and antral follicles in ovaries compared with cattle with a high AFC (Ireland *et al.* 2008).









#### Hormones

Transient rises in FSH concentrations precede and stimulate the emergence of follicle waves (Adams et al. 1992). A number of studies have now shown that FSH concentrations are lower in animals with high AFC compared with animals with low AFC (Burns et al. 2005, Ireland et al. 2007, Mossa et al. 2010b). In addition, circulating LH concentrations during the estrous cycle are chronically higher in young adult cattle with low versus high AFC (Jimenez-Krassel et al. 2009). This observation leads to the question as to whether pituitary function differs between the two groups of animals. We tested this notion by examining the response to ovariectomy of cows with high or low AFC and then challenging them with GnRH and follicular fluid (rich source of inhibin) on separate occasions 30 days after ovariectomy (Mossa et al. 2010b). The study showed that there was no difference between the two groups in basal FSH or LH secretion or response to the treatments leading to the conclusion that differences in gonadotrophin secretion between cows with high versus low AFC is due to differences in negative feedback hormones and not differences in pituitary function (Mossa et al. 2010b).

Evidence that the total number of healthy follicles in ovaries contributes to the endocrine environment is provided by examining Anti-Müllerian Hormone (AMH) concentrations. AMH is produced primarily by granulosa cells of healthy growing follicles (La Marca & Volpe 2006) and is positively associated with follicle numbers in mice (Kevenaar et al. 2006) and women (Jayaprakasan et al. 2009, van Disseldorp et al. 2009). It has recently been shown that AMH concentrations can reliably predict the AFC and the number of healthy follicles and oocytes in age matched young adult cattle (Ireland et al. 2008). Furthermore, AMH serum concentrations before superovulation are highly positively correlated with the numbers of ovulations after treatment (Rico et al. 2009) and are repeatable within animals (Ireland et al. 2008, Rico et al. 2009). In comparison to AMH, serum concentrations of both oestradiol and inhibin-A, the main feedback regulators of FSH secretion, are not different between groups of animals with high and low AFC (Burns et al. 2005, Ireland et al. 2007). This may be explained by the fact that individual follicles from cattle with low AFC have higher follicular fluid concentrations and produce more oestradiol than follicles from animals with high AFC (Ireland et al. 2009) and we have speculated that this difference may be a consequence of higher circulating FSH concentrations in the low versus the high AFC groups (Ireland et al. 2009). Nonetheless it appears that the total oestradiol negative feedback is similar between the high and low groups as fewer follicles produce more oestradiol in the low AFC animals compared with more follicles producing lower oestradiol in the high AFC group (at least within the limits of sensitivity of our oestradiol assay). Despite not being able to measure a difference in circulating oestradiol concentrations between high and low AFC groups, we have recently shown that circulating androgen concentrations are indeed higher in cattle with high compared with low AFC (Jimenez-Krassel et al. 2009). In the absence of an alternative explanation, we suggest that the difference in circulating gonadotrophin concentrations associated with variable AFC is due to differences in ovarian feedback hormones produced by the follicles.

Due to the considerable interest in the contribution of early/mid luteal phase progesterone concentrations for the successful establishment of pregnancy, we compared progesterone concentrations between groups of animals and found that animals with low AFC have much lower progesterone concentrations during oestrous cycles than animals with high AFC (Jimenez-Krassel et al. 2009) (Figure 3). Also the ability of both granulosa and luteal cells from animals with high AFC to produce progesterone in vitro was diminished compared with animals with high AFC (Jimenez-Krassel et al. 2009). The possible consequences of this difference in progesterone concentrations are discussed below.



**Fig. 3.** Alterations in serum concentrations of progesterone during the bovine oestrous cycle (Day 0 = oestrus). Each symbol represents the daily mean ( $\pm$ SEM) progesterone value for animals with a consistently low ( $\leq$  15 follicles per wave, n = 32 oestrous cycles for 25 animals) versus high ( $\geq$  25 follicles per wave, n = 30 cycles for 22 animals) AFC during follicular waves. Asterisks indicate significant differences (\*\* P < 0.01) between groups. ANOVA indicated an overall significant effect of group (Low versus High, P<0.02), day of cycle (P<0.001) and group x day interaction (P<0.001). From (Jimenez-Krassel *et al.* 2009).

# Causes of the variation in the numbers of ovarian follicles

The cause of the inherently high variation in number of follicles is unknown but a number of explanations are plausible. Firstly, proliferation of oogonia during early embryonic development (that give rise to a fixed pool of non proliferating oocytes within primordial follicles for the lifetime of the animal) may be highly variable resulting in a highly variable pool of follicles. Secondly, proliferation of oogonia may be relatively constant (similar among females) but depletion of this pool may proceed at different rates among individuals giving rise to females with differing AFC at the same age. Thirdly, the variation in follicle numbers among age matched females occurs as a result of a combination of the first two possibilities.

David Barker (the Barker Hypothesis) suggests that environmental influences early in human foetal life are reflected in impaired growth, development and metabolism leading to increased risk for diseases in adulthood (Barker 1992). The hypothesis proposes that some diseases originate through foetal adaptations to malnutrition that permanently alter body function. This hypothesis is supported by animal models (McMillen & Robinson 2005). Most studies have examined the link between malnutrition with cardiovascular disease, obesity and diabetes, but few studies have examined potential links with reproduction. However, poor foetal growth due to the manipulation of maternal diet alters gonadotrophin gene expression in the pituitary and number of follicles in the ovaries of late-gestation lambs (Da Silva *et al.* 2002). In addition, maternal under-nutrition during the first trimester retards foetal ovarian development in sheep (Borwick *et al.* 1997). Hence, it is reasonable to speculate that maternal nutrition during gestation, at the time of ovarian development in their foetuses, may impact oogonia proliferation and thus follicle numbers postnatally. We have recently started to test this hypothesis by restricting nutrition of beef heifers to 0.6 of their maintenance energy requirements, from shortly before

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conception to the end of the first trimester of pregnancy (period encompassing the peak in oocyte numbers in foetuses (Erickson 1966a)). Results show that calves born to nutritionally restricted mothers have a 60% lower peak, minimum and mean AFC during follicular waves compared with calves born to mothers fed control diets (Mossa *et al.* 2009) (Table 1). In addition, calves born to dams nutritionally restricted also show higher resting arterial blood pressure compared with those born to control mothers (Mossa *et al.* 2009). In addition, similar studies in *Bos indicus* cross heifers support the notion that prenatal maternal nutrition affects ovarian measures in their offspring (Sullivan *et al.* 2009).

Heifer age (weeks)	Antral Follicle Count		
	Control $(n = 13)$	Restricted $(n = 10)$	P<
7	$23.9 \pm 2.1$	$14.1\pm0.9$	0.01
18	$26.1 \pm 2.9$	$16.2 \pm 1.1$	0.01
35	$23.9 \pm 2.2$	$16.6 \pm 1.2$	0.01

 Table 1. Mean (± SEM) number of follicles ≥3 mm in diameter (Antral Follicle Count) during follicle waves in heifer calves born to mothers fed a control diet (Control) or who were nutritionally restricted (Restricted, 60% energy requirement) for the first 110 days of gestation. From (Mossa et al. 2009).

The Barker Hypothesis, and supporting data in a variety of animal models, indicates that a negative uterine environment can influence the development of different organs and systems in the foetus including the reproductive system. This observation may have relevance for the dairy industry, since it is well established that selecting dairy cows for increased milk production potential has been associated with a concomitant decrease in fertility and an increase in the susceptibility to some diseases (Beckers et al. 2002). In order to have a 365-day calving interval, cows must conceive during the period of their peak lactation. This period of peak metabolic stress is coincident with follicle growth, ovulation, fertilization, early embryonic development and early foetal development. According to the Barker Hypothesis, animals conceived and developing in this nutritionally stressed maternal environment may have compromised development that could affect them for the rest of their lives. We have investigated the relationship between maternal milk production (a proxy for maternal stress) on performance indicators in the first, second, and third lactations of their female offspring using a large Irish database and results have shown that greater milk yield preconception and during gestation is associated with reduced survival and milk yield and greater somatic cell count in their progeny (Berry et al. 2008). This shows that maternal environment during gestation in dairy cows does have longterm effects on their female offspring but we have not yet investigated the effects of AFC.

## Consequences of the variation in the numbers of ovarian follicles

In women undergoing IVF treatment, the number of antral follicles present before ovarian stimulation is considered a predictor of the ovarian responsiveness to gonadotrophin stimulation, since low numbers of follicles are associated with lower numbers of oocytes recovered and pregnancy rates (Tomas *et al.* 1997, Chang *et al.* 1998, Hsieh *et al.* 2001, Huang *et al.* 2001, Beckers *et al.* 2002). Similarly in cattle the number of follicles prior to superovulatory treatment is correlated with the numbers of follicles and corpora lutea after superovulation, and total ova and transferable embryos recovered (Kawamata 1994, Cushman *et al.* 1999, Taneja *et al.* 2000, Singh *et al.* 2004). Moreover, intrafollicular oestradiol concentrations are about 2-fold higher in animals with a low versus a high AFC (Ireland *et al.* 2009) and this may have detrimental effects on oocyte maturation and developmental competence in cattle with low follicle

#### Variation in numbers of ovarian follicles

numbers because high physiological concentrations of oestradiol block maturation of bovine oocytes in vitro (Beker-van Woudenberg *et al.* 2004). In addition, lower circulating concentrations of progesterone in cattle with low AFC are associated with a much lower endometrial thickness from Day 0 to 6 of the oestrous cycle (Jimenez-Krassel *et al.* 2009) and endometrial thickness is positively associated with implantation and pregnancy rates in humans subjected to in vitro fertilization programs (Raga *et al.* 1999). Taken together these findings indicate that AFC is positively associated with fertility in cattle. To test this hypothesis we performed ovarian ultrasonography on 306 dairy cows (aged  $3.48 \pm 1.66$  years) during the first wave of follicular development 1 to 4 months post-partum and recorded their reproductive performances during the breeding season. Cows with a high AFC had higher pregnancy rates, shorter calving to conception intervals and received fewer services during the breeding season compared with cows with a low AFC (Mossa *et al.* 2010a). This is supported by a study in beef heifers showing higher pregnancy rates in heifers with high AFC versus low AFC (Cushman *et al.* 2009).

## Conclusion

Improved understanding of the pattern of follicle development in the last 20 years has led to the development of more effective treatments to synchronize (and induce) oestrus behaviour in cattle. The recent studies reviewed here indicate that the variation in number of follicles growing during follicle waves, and in toto in ovaries, may be an important consideration when new methods are developed to manipulate and improve superovulation and fertility in cattle. Moreover, the variation in number of oocytes and follicles in offspring may be determined by the maternal environment during foetal development coincident with development of all the organ systems. Thus, the high variation in follicle numbers in adults may not only be reflective of reproductive disorders and suboptimal fertility, but also alterations in other non-reproductive systems that may have profound effects on the animal's health and welfare.

## Grant support

AE, FM, TF and PL are supported by grants from Science Foundation Ireland (07/SRC/B1156) and from the Irish Department of Agriculture, Fisheries and Food, Research Stimulus Fund programme 2006 (RSF 06-328).

SB is supported by funds from the National Development Plan, the Irish Dairy Levy Trust, and Irish Department of Agriculture, Fisheries and Food, Research Stimulus Fund programme 2006 (RSF 06-328).

AEZ-S is supported by a grant from the Polish Ministry of Science and Higher Education (N N311 324136).

GWS is supported by grants from the National Research Initiative Competitive Grant no. 2005-35203-16011 from the USDA National Institute of Food and Agriculture, Agriculture and Food Research Initiative Competitive Grant no. 2009-65203-05700 from the USDA National Institute of Food and Agriculture, and by the Michigan Agricultural Experiment Station

JJI is supported by National Research Initiative Competitive Grants 2001-002255, 2004-01697 and 2007-01289 from the USDA Cooperative State Research, Education and Extension Service and by funds from the Michigan Agriculture Experiment Station.

#### Acknowledgements

We thank D Berry for his comments on this manuscript.

## References

- Adams GP, Matteri RL, Kastelic JP, Ko JCH & Ginther OJ 1992 Association between surges of folliclestimulating hormone and the emergence of follicular waves in heifers. *Journal of Reproduction and Fertility* 94 177-188.
- Barker DJ 1992 The effect of nutrition of the fetus and neonate on cardiovascular disease in adult life. *Proceedings of the Nutrition Society* **51** 135-144.
- Beckers NGM, Macklon NS, Eijkemans MJC & Fauser BCJM 2002 Women with regular menstrual cycles and a poor response to ovarian hyperstimulation for in vitro fertilization exhibit follicular phase characteristics suggestive of ovarian aging. *Fertility and Sterility* 78 291-297.
- Beker-van Woudenberg AR, van Tol HTA, Roelen BAJ, Colenbrander B & Bevers MM 2004 Estradiol and its membrane-impermeable conjugate (estradiol-bovine serum albumin) during in vitro maturation of bovine oocytes: effects on nuclear and cytoplasmic maturation, cytoskeleton, and embryo quality. *Biol Reprod* **70** 1465-1474.
- Berry DP, Lonergan P, Butler ST, Cromie AR, Fair T, Mossa F & Evans ACO 2008 Negative influence of high maternal milk production before and after conception on offspring survival and milk production in dairy cattle. Journal of Dairy Science **91** 329-337.
- Borwick SC, Rhind SM, McMillen SR & Racey PA 1997 Effect of undernutrition of ewes from the time of mating on fetal ovarian development in mid gestation. *Reproduction Fertility and Development* **9** 711-715.
- Burns DS, Jimenez-Krassel F, Ireland JL, Knight PG & Ireland JJ 2005 Numbers of antral follicles during follicular waves in cattle: evidence for high variation among animals, very high repeatability in individuals, and an inverse association with serum follicle-stimulating hormone concentrations. *Biology* of *Reproduction* **73** 54-62.
- Chang MY, Chiang CH, Hsieh TT, Soong YK & Hsu KH 1998 Use of the antral follicle count to predict the outcome of assisted reproductive technologies. *Fertility and Sterility* **69** 505-510.
- Cushman RA, Allan MF, Kuehn LA, Snelling WM, Cupp AS & Freetly HC 2009 Evaluation of antral follicle count and ovarian morphology in crossbred beef cows: investigation of influence of stage of the estrous cycle, age, and birth weight. *Journal of Animal Science* 87 1971-1980.
- Cushman RA, DeSouza JC, Hedgpeth VS & Britt JH 1999 Superovulatory response of one ovary is related to the micro- and macroscopic population of follicles in the contralateral ovary of the cow. *Biology of Reproduction* **60** 349-354.
- Da Silva P, Aitken RP, Rhind SM, Racey PA & Wallace JM 2002 Impact of maternal nutrition during pregnancy on pituitary gonadotrophin gene expression and ovarian development in growth-restricted and normally grown late gestation sheep fetuses. *Reproduction* 123 769-777.

- Erickson BH 1966a Development and radio-response of the prenatal bovine ovary. *Journal of Reproduction and Fertility* **11** 97-105.
- Erickson BH 1966b Development and senescence of the postnatal bovine ovary. *Journal of Animal Science* 25 800-805.
- Evans ACO 2003 Characteristics of ovarian follicle development in domestic animals. *Reproduction in Domestic Animals* 38 1-7.
- Hsieh Y-Y, Chang C-C & Tsai H-D 2001 Antral follicle counting in predicting the retrieved oocyte number after ovarian hyperstimulation. *Journal of Assisted Reproduction and Genetics* **18** 320-324.
- Huang F-J, Chang S-Y, Tsai M-Y, Kung F-T, Wu J-F & Chang H-W 2001 Determination of the efficiency of controlled ovarian hyperstimulation in the gonadotropin-releasing hormone agonist-suppression cycle using the initial follicle count during gonadotropin stimulation. *Journal of Assisted Reproduction and Genetics* 18 91-96.
- Ireland JJ, Mihm M, Austin E, Diskin MG & Roche JF 2000 Historical perspective of turnover of dominant follicles during the bovine estrous cycle: key concepts, studies, advancements, and terms. *Journal of Dairy Science* **83** 1648-1658.
- Ireland JJ & Roche JF 1982 Development of antral follicles in cattle after prostaglandin-induced luteolysis: changes in serum hormones, steroids in follicular fluid, and gonadotropin receptors. *Endocrinology* 111 2077-2086.
- Ireland JJ & Roche JF 1983a Development of nonovulatory antral follicles in heifers: changes in steroids in follicular fluid and receptors for gonadotropins. *Endocrinology* **112** 150-156.
- Ireland JJ & Roche JF 1983b Growth and differentiation of large antral follicles after spontaneous luteolysis in heifers: changes in concentrations of hormones in follicular fluid and specific binding of gonadotropins to follicles. *Journal of Animal Science* **57** 157-167.
- Ireland JJ & Roche JF 1987. Hypotheses regarding development of dominant follicles during a bovine estrous cycle. In Follicular Growth and Ovulation Rate in Farm Animals, pp. 1-18.
- Ireland JJ, Ward F, Jimenez-Krassel F, Ireland JL, Smith GW, Lonergan P & Evans ACO 2007 Follicle numbers are highly repeatable within individual animals but are inversely correlated with FSH concentrations and the proportion of good-quality embryos after ovarian stimulation in cattle. *Human Reproduction* 22 1687-1695.
- Ireland JJ, Zielak-Steciwko AE, Jimenez-Krassel F, Folger J, Bettegowda A, Scheetz D, Walsh S, Mossa F, Knight PG, Smith GW, Lonergan P & Evans ACO 2009 Variation in the ovarian reserve is linked to alterations in intrafollicular estradiol production and ovarian biomarkers of follicular differentiation and oocyte quality in cattle. *Biology of Reproduction* 80 954-964.

Ireland JLH, Scheetz D, Jimenez-Krassel F, Themmen

AP, Ward F, Lonergan P, Smith GW, Perez GI, Evans AC & Ireland JJ 2008 Antral follicle count reliably predicts number of morphologically healthy oocytes and follicles in ovaries of young adult cattle. *Biology of Reproduction* **79** 1219-1225.

- Jayaprakasan K, Deb S, Batcha M, Hopkisson J, Johnson I, Campbell B & Raine-Fenning N 2009 The cohort of antral follicles measuring 2-6 mm reflects the quantitative status of ovarian reserve as assessed by serum levels of anti-Mullerian hormone and response to controlled ovarian stimulation. *Fertility and Sterility* In Press 2009 Nov 2018. [Epub ahead of print].
- Jimenez-Krassel F, Folger JK, Ireland JL, Smith GW, Hou X, Davis JS, Lonergan P, Evans ACO & Ireland JJ 2009 Evidence that High Variation in Ovarian Reserves of Healthy Young Adults Has a Negative Impact on the Corpus Luteum and Endometrium During Estrous Cycles in Cattle. *Biology of Reproduction* **80** 1272-1281.
- Kawamata M 1994 Relationship between the number of small follicles prior to superovulatory treatment and superovulatory response in Holstein cows. *Journal of Veterinary Medical Science* 56 965-970.
- Kevenaar ME, Meerasahib MF, Kramer P, van de Lang-Born BM, de Jong FH, Groome NP, Themmen AP & Visser JA 2006 Serum anti-mullerian hormone levels reflect the size of the primordial follicle pool in mice. Endocrinology 147 3228-3234.
- La Marca A & Volpe A 2006 Anti-Mullerian hormone (AMH) in female reproduction: is measurement of circulating AMH a useful tool? *Clinical Endocrinol*ogy **64** 603-610.
- Lussier JG, Matton P & Dufour JJ 1987 Growth rates of follicles in the ovary of the cow. *Journal of Reproduction and Fertility* 81 301-307.
- McMillen IC & Robinson JS 2005 Developmental origins of the metabolic syndrome: prediction, plasticity, and programming. *Physiological Reviews* 85 571-633.
- Mihm M, Crowe MA, Knight PG & Austin EJ 2002 Follicle wave growth in cattle. *Reproduction in Domestic Animals* 37 191-200.
- Mossa F, Butler ST, Berry DP, Jimenez-Krassel F, Lonergan P, Smith GW, Ireland JJ & Evans ACO 2010a. Number of follicles during follicular waves is positively associated with fertility in dairy cows. In *Reproduction in Domestic Ruminants VII*, in this volume.
- Mossa F, Jimenez-Krassel F, Walsh S, Berry DP, Butler ST, Folger J, Smith GW, Ireland JL, Lonergan P, Ireland JJ & Evans AC 2010b Inherent capacity of the pituitary gland to produce gonadotropins is not influenced by the number of ovarian follicles > or = 3 mm in diameter in cattle. *Reproduction, Fertility* and Development **22** 550-557.
- Mossa F, Kenny D, Jimenez-Krassel F, Smith GW, Berry D, Butler S, Fair T, Lonergan P, Ireland JJ & Evans

ACO 2009 Undernutrition of heifers during the first trimester of pregnancy diminishes size of the ovarian reserve in female offspring. *Biology of Reproduction*, **81** (Supplement 1), 135.

- Pierson RA & Ginther OJ 1984 Ultrasonography of the bovine ovary. Theriogenology 21 495-504.
- Pierson RA & Ginther OJ 1988 Ultrasonic imaging of the ovaries and uterus in cattle. *Theriogenology* 29 21-37.
- Raga F, Bonilla-Musoles F, Casan EM, Klein O & Bonilla F 1999 Assessment of endometrial volume by threedimensional ultrasound prior to embryo transfer: clues to endometrial receptivity. *Human Reproduction* 14 2851-2854.
- Rajakoski E 1960 The ovarian follicular system in sexually mature heifers with special reference to seasonally, cyclical, and left-right variations. *Acta Endocrinologica* Supplement 52 1-68.
- Rico C, Fabre S, Medigue C, di Clemente N, Clement F, Bontoux M, Touze JL, Dupont M, Briant E, Remy B, Beckers JF & Monniaux D 2009 Anti-mullerian hormone is an endocrine marker of ovarian gonadotropin-responsive follicles and can help to predict superovulatory responses in the cow. *Biology of Reproduction* 80 50-59.
- Savio JD, Keenan L, Boland MP & Roche JF 1988 Pattern of growth of dominant follicles during the oestrous cycle of heifers. *Journal of Reproduction and Fertility* 83 663-671.
- Singh J, Dominguez M, Jaiswal R & Adams GP 2004 A simple ultrasound test to predict the superstimulatory response in cattle. *Theriogenology* **62** 227-243.
- Sirois J & Fortune JE 1988 Ovarian follicular dynamics during the estrous cycle in heifers monitored by real-time ultrasonography. *Biology of Reproduction* 39 308-317.
- Sullivan TM, Micke GC, Greer RM, Irving-Rodgers HF, Rodgers RJ & Perry VE 2009 Dietary manipulation of Bos indicus x heifers during gestation affects the reproductive development of their heifer calves. *Reproduction, Fertility and Development* **21** 773-784.
- Taneja M, Bols PE, Van de Velde A, Ju JC, Schreiber D, Tripp MW, Levine H, Echelard Y, Riesen J & Yang X 2000 Developmental competence of juvenile calf oocytes in vitro and in vivo: influence of donor animal variation and repeated gonadotropin stimulation. *Biology of Reproduction* 62 206-213.
- Tomas C, Nuojua-Huttunen S & Martikainen H 1997 Pretreatment transvaginal ultrasound examination predicts ovarian responsiveness to gonadotropins in in-vitro fertilization. *Human Reproduction* **12** 220-223.
- van Disseldorp J, Lambalk CB, Kwee J, Looman CW, Eijkemans MJ, Fauser BC & Broekmans FJ 2009 Comparison of inter- and intra-cycle variability of anti-Mullerian hormone and antral follicle counts. *Human Reproduction* 25 221-227.